Practical and versatile products for hygiene control and environmental monitoring





NRS™ Transwab®

Environmental monitoring of disinfected surfaces

If swab samples are taken from surfaces which have been disinfected, there may still be traces of biocide which can react with and reduce the count of microorganisms present at the time of sampling. Such microorganisms may have been part of, and protected by a surface biofilm, and the action of swabbing loosens them and exposes them to the biocide. This will result in a false low count, and failure to detect a potential hazard for subsequent food or beverage production.

It is essential, therefore, and may be mandatory, that any residual disinfectant on surfaces is neutralised at the time of sampling by first wetting the swab in an appropriate neutralising solution.

NRS™ Medium contains lecithin, polysorbate 80, and sodium thiosulphate, in a peptone* phosphate buffer¹². This formulation is in accordance with ISO 18593:2004¹, and will neutralise most disinfectants used in the food industry, including those based on chlorine, peroxygen compounds, iodine, quaternary ammonium compounds, amphoterics, biguanides, and glutaraldehyde¹.

NRS™ Transwabs® are supplied with a transport tube containing precisely 5ml or 10ml of NRS™ medium to allow accurate quantitative assessment of contamination levels.

NRS Transwabs can also be used for swabbing in slaughterhouses and meat cutting plants in compliance with ISO 17604:2003 and European Commission Regulation (EC) No. 2073/2005 (Non-destructive methods) for the "Wet and Dry Swab Method" (ISO 17604), our Steriswab Dry can be used as the dry swab component.

- Certificate of conformity supplied with each pack
- Manufactured to ISO 9001:2000
- Conforms to ISO 18593:2004
- Conforms to European Commission Regulation (EC) No. 2073/2005
- Conforms to requirements of ISO 17604:2003
- Forms part of HACCP and GMP programmes
- Guaranteed 2% fill tolerance
- Isotonic NRSTM neutralises residual disinfectants and sanitisers

*Peptone in MW& E products is of vegetable origin

Technique:

Swabs are moistened in the neutralising buffer before swabbing the specified area, normally 100cm², of the test surface. The swab is then placed in the Transwab³ containing either 5ml or 10ml of NRS¹¾. Complete the tube label and transport to the laboratory. The swab and/or buffer can be plated directly onto a suitable growth medium or the NRS¹¾ buffer can be membrane-filtered to concentrate any microorganisms present.



Steriswab™

Steriswabs[™] have been designed for the pharmaceutical, industrial and food microbiologist as a tool for effective monitoring of sterility and contamination in clinically clean or sterile work areas.

Steriswabs^{TX} are pre-moistened with a sterile diluent for ease of use and the elimination of any possible cross-contamination. The swabs are supplied individually in a sterile labelled tube which can be used to return the sample to the laboratory.

SteriswabsTM are pre-sterilised by irradiation and triple-wrapped to guarantee that any viable organisms detected come from the site and not from the swab. Our own production procedures are designed to ensure minimum bioburden (i.e. practically zero non-viables).

SteriswabsTM are validated for the recovery of fastidious microorganisms.

Steriswab[™] dry

Made to the same rigorous standard as the pre-moistened Steriswab^{ne}, but kept dry to allow use with customer's own diluent, or can be used dry if required.

Dryswab™

A versatile range of swabs for all your sampling requirements.

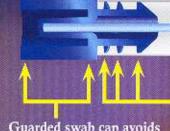








Security label guarantees sterility and integrity



Guarded swab cap avoids operator contamination Fluted cap ensur containment of s triple security



Polywipes"

Unique sponge for environmental sampling

Pre-moistened 50cm2 blue sponge swabs for easy and effective sampling for the detection of pathogens such as Listeria.

The bright blue colour of the sponge material is highly visible, to ensure easy detection and removal if it should be dropped in a work area. PolywipesTM are prepared using a specially manufactured sponge material designed to have no inhibitory substances, and to ensure maximum



GUARANTEED recovery and optimum RECOVERY detection of all micro-organisms.

Conventional sponge materials contain inhibitory substances such as orthophenylphenol, sulphur and quaternary ammonium compounds, either by design, or left by the manufacturing process. Some of these chemicals are also used in the food industry for sanitation. These can kill or damage any microorganisms picked up and therefore prevent their recovery and detection in the laboratory ...

Polywipes™ with drawcord

Pre-moistened sponge swabs complete with integral 1.2m drawcord for easy and effective sampling of floor spaces for the effective detection of pathogens.

The method was originally developed for the detection of salmonellae in poultry farms, being particularly good for the sampling of litter and dropping surfaces in chicken houses, but is also suitable for other industrial monitoring applications*.9.

The drawcord can be drawn by hand, or attached to to clothing and dragged behind while walking to allow a large area to be thoroughly sampled. The method is ideal for floor areas such as poultry pens, food processing factories, bakeries etc. It is also useful for sampling inaccessible areas such as vats and drains.

Polywipes™ with drawcord are packed in conventional peel pouches. They are supplied on their own, or complete with resealable polythene bags for transport of the sample to the laboratory. Sterile gloves can also be supplied if required.

Polywipes SKTB

PolywipesTM with drawcord & premoistened with skimmed milk.

Some users require the use of draggable sponge swabs premoistened with skimmed milk. This has been reported to improve the survival of Salmonellae at reduced temperatures 10.11

Polywipes™ with Peptone Saline for Carcass Sampling

Recent regulations require the sampling of carcasses of cattle, sheep or pigs using sponge swabs and according to certain procedures. Polywipes™ with Peptone Saline (MW726) are particularly suitable for this 6,1226

Procedure (for EU**)

Use PolywipesTM sponge lengthwise.

Sample carcass according to prescribed procedure for species. Estimate surface area swabbed by multiplying approximate length of carcass in cm by the width of the Polywipes™ sponge (10cm), e.g. Swabbing I metre (100cm) of pig carcass surface with sponge with width 10cm gives swabbed area of 100cm x 10cm = 1000cm2,

Place PolywipesTM in sample bag.

Label bag indicating carcass reference, farm (with postcode)species, area swabbed, and send sample to laboratory.

**In non-EU countries please refer to national authorities for details of any statutory procedures.

Flexiswab**

The longer flexible-shafted swab

Strong flexible longer 8 inch/205 mm shaft for extra reach around bends without breaking.

The blue shaft for maximum visibility has a larger bud for effective sampling with non toxic properties and good absorption.

- Tamper-evident seal ensures sterility before sampling
- Labelled tube for site details and transport of sample
- Ideal for taps, pipes, vents, valves, vats, machinery, crevices and other awkward sites

mple with

5ml or 10ml variants available with critical fill control NRSTM guarantees recovery of microorganisms

Rayon/viscose swab enhances the recovery of microorganisms and provides both good absorption and release of sample



Order information



Code	Description Qua	
MW 774	Sterile blue shaft rayon swap (with breakpoint) and tube with 5ml NRS	100
MW 775	Sterile blue shaft rayon swab (with breakpoint) and tube with 10ml NRS	100

Flexiswab™ & Dryswab™

Code	Description	Quantity
MW 159	Flexible 8 nch/205mm edra long blue shaft, rayon, tubed, sterile	100
MW 102	Plastic shaft, rayon, tubed, sterile	800
MW 103	Plastic shaft, alginate, tubec, sterile	100
MW 104	Wood shaft, rayon, tubed, sterle	800
MW 104J	Wood shaft, extra large rayon, tubed, sterile	100
MW 107	Wood shaft, alginate, tubed, sterile	100
MW 108	Wood shaft, rayon, peel pouch, sterila	1250
MW 112	Plastic shaft rayon, peel pouch, sterile	1250
MW 116	Wood shaft, rayon, bulk packed (50 x 100), non-sterile	5000
MW 116S	Wood shaft, rayon, bulk packed (50 x 100), sterile	5000
MW 118	Pastic shaft, rayon, bulk packed (50 x 100), non-sterile	5000
MW 118S	Plastic shaft, rayon, bulk packed (50 x 100), sterile	5000



Code	Description Quan	tity
MW726	Polywipe" premoistened with Peptone Saline (for carcasses)	50
MW727	Polywipe" premaistened with 1.2m* ntegral drawcord in pee pouch	50
MW727A	Polywipe or premaistened with 1.2m integral drawcord in peer pouch, with 50 resealable bags.	50
MW727SKTB	Polywipe" premoistened with skinimed milk medium with 1.2m integral drawcord in peel pouch with 50 twist bags.	50
MW728	Polywipe" premoistened in 100ml container and gloves	30
MW729	Polywipe" premoistened with peel pouch	50
MW729A	Polywice" premoistened with peel pouch and 50 researche bags	50
MW729B	Polywipe" premosteried with peel pouch, 50 resealable bags and sterile gloves	50
MW729TB	Polywipe" premostened with poel pouch, 50 twist bags and sterile gloves:	50
*romnal leng	pth of crawcord	

Code	Description	uantity
MW 720	Pre-moistened rayon swab (breakpoint), labelled tube, triple wrapped	250
MW 730	Dry rayon swab (breakpoint), labelled tube, triple wrapped	250
MW 735/1	Dry rayon swab (breakpoint), peel bouch	150
MW 735/5	Dry rayon swab (oreakpoint), peel pouch, five per pouch (200x 5)	1000
MW 735/10	Dry rayon swab (breakpoint), peel pouch, ten per pouch (100x 10)	1000

References

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- 3 ISO 17604:2003 Microbiology of food and animal feeding stuffs-Carcass sampling for microbiological analysis. International Standards Organisation, Geneva
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- 5 Silliker, J.H., & D.A. Gabis, 1975, A Cellulose Sponge Sampling Technique for Surfaces, J. Milk Food Technol. 38, 504
- 6 Daley, E.F., F. Paganto & J.M.Farber, 1995. The inhibitory properties of various sponges on Listeria spp. Letters in Applied Microbiology, 20, 195-198
- Gordon, L., C. Papelou, K. Pathak, & S. Surman, 2002, A Preliminary Study To Assess Inhibitory Effects of Different Types of Sponge Material on Microorganisms, Poster P12, Public Health Laboratory Service, 27th Annual Scientific Conference, Warwick
- 8 Kingston, D.J., 1981, A Comparison of Culturing Drag Swabs and Litter for Identification of Infections with Salmonella spp. In Commercial Chicken Flocks, Avian Diseases 25:513-516
- 9 Mallinson, E.T., C.R Tate, R.G.Miller, B.Bennett, & E. Russek-Cohen, 1989, Monitoring Poultry Farms for Salmonella by Drag Swab Sampling and Antigen-Capture Immunoassay, Avian Diseases 33:684-689

- 10 Mallinson, E.T., 1991, Novel Salmonella Detection System Developed: Combines Reliability, Practicality, Feedstuffs 63:40-44
- 11 Opara, O.O., et al., 1992. The Effect of Exposure, Storage Times, and Types of Holding Media on the Drag-Swah Monitoring Technique for Salmonella, Avian Diseases 36:63-68
- 12 Statutory Instrument 2006 No. 14 The Food Hygiene (England) Regulations 2006
- 13 The Food Hygiene (Scotland) Regulations 2006 (SSI 2006/3)
- 14 The Food Hygiene (Wales) Regulations 2006 (SI 2006/31 (W.5)
- 15 The Food Hygiene Regulations (Northern Ireland) 2006 (SR 2006 No 3)
- 16 Food Standards Agency, Guide to Food Hygiene & Other Regulations for the UK Meat Industry, www.food.gov.uk/foodindustry/meat/guidehygienemeat



ISO 9001



Steriswab"